

SUPPLEMENTARY APPENDIX

Tezacaftor-Ivacaftor in Patients with Cystic Fibrosis and Phe508del and a Residual Function Mutation

*Steven M. Rowe, MD, MSPH, Cori Daines, MD, Felix C. Ringshausen, MD, Eitan Kerem, MD, John Wilson, FRACP, PhD, Elizabeth Tullis, MD, FRCPC, Nitin Nair, PhD, Christopher Simard, MD, Linda Han, MD, MPH, Edward P. Ingenito, MD, Charlotte McKee, MD, Julie Lekstrom-Himes, MD, *Jane C. Davies, MD, MBChB, FRCPCH

**These authors contributed equally to this work*

TABLE OF CONTENTS

LIST OF INVESTIGATORS.....	3
SUPPLEMENTAL METHODS.....	6
Criteria for Eligible Residual Function Mutations	6
Prespecified Patient-Level Eligibility Criteria	6
Sample Size and Power	7
Analysis of Primary Endpoint Variables	7
Mixed-Model for Repeated Measures Analysis of Primary Endpoint	8
Subgroup Analysis of the Primary Efficacy Endpoint	9
Analysis of Key Secondary Efficacy Endpoint	10
Other Secondary Endpoints.....	10
Multiplicity Adjustment.....	11
SUPPLEMENTAL RESULTS	11
Exploratory and Additional Supportive Endpoints	11
Safety	13

Additional Information on Adverse Events	13
Respiratory Adverse Events	13
Acute Effects on Spirometry	14
Liver Function Tests	14
SUPPLEMENTAL DISCUSSION	15
SUPPLEMENTAL REFERENCES	16
SUPPLEMENTAL TABLES	17
Table S1. Eligible Residual Function Mutations	17
Table S2. Summary of Treatment-Emergent Respiratory Events by Preferred Term, Safety Set.	18
Table S3. Mean (SD) Change from Predose to Post Dose in Percentage of Predicted FEV ₁ on Days 1 and 15, Safety Set for Patients ≥12 to <18 Years Old at Screening.	19
Table S4. Liver Transaminase and Bilirubin Elevations During the Study Period.	20
SUPPLEMENTAL FIGURES	21
Figure S1. Study Design.	21
Figure S2. CONSORT Diagram.	22

LIST OF INVESTIGATORS

The VX14-661-108 study group included: Steven M. Rowe, University of Alabama at Birmingham; Noah Lechtzin, The Johns Hopkins Hospital; Richard C. Ahrens, The University of Iowa Hospitals and Clinics; Scott H. Donaldson, UNC Marsico Clinical Research Center; Kimberley Ann McBennett; University Hospitals Cleveland Medical Center/Rainbow Babies and Children's Hospital; Joseph M. Pilewski, Children's Hospital of Pittsburgh of University of Pittsburgh Medical Center; Theresa Laguna, University of Minnesota; Carlos Milla, Stanford University; Ronald Rubenstein, Children's Hospital of Philadelphia; Daniel B. Rosenbluth, Washington University School of Medicine / St. Louis Children's Hospital; Seth Walker, The Emory Clinic / Children's Healthcare of Atlanta at Egleston; Michael R. Powers, Oregon Health & Science University; Christopher N. Fortner, SUNY Upstate Medical University; Theodore G. Liou, University of Utah / Primary Children's Medical Center; Maria Berdella, Beth Israel Medical Center; Patricia Joseph, UC Health Clinical Trials Office; Lara Bilodeau, Institut Universitaire de Cardiologie et Pneumologie de Quebec; Manu Jain, Northwestern Memorial Hospital; Samya Nasr, University of Michigan Health System; Jennifer Taylor-Cousar, National Jewish Health; Diana Quintero, Children's Hospital of Wisconsin; Bryon Quick, Kaiser Permanente; Ahmet Uluer, Boston Children's Hospital; Bruce A. Barnett, The Toledo Hospital/Toledo Children's Hospital; Emily DiMango, Columbia University Medical Center; Patrick Flume, Medical University of South Carolina; Floyd Livingston, Nemours Research Institute; Gregory J. Omlor, Akron Children's Hospital; Alix Ashare, Dartmouth Hitchcock Medical Center, Lebanon; Matthias Salathe, University of Miami/Miller School of Medicine; Allen Dozor, New York Medical College; Barbara Messori, Azienda

Ospedaliero Universitaria San Luigi Gonzaga; Edith Zemanick, Children's Hospital
 Colorado; Raksha Jain, The University of Texas Southwestern Medical Center; Michael
 McCarthy, Providence Pediatric Pulmonary & Allergy/Immunology Clinic; Tarik Haddad,
 Tampa General Hospital Cardiac and Lung Transplant Clinic; Julie Philley, The
 University of Texas Health Center at Tyler; Daniel T. Layish, Central Florida Pulmonary
 Group; Terry W. Chin, Miller Children's Hospital / Long Beach Memorial; Cori Daines,
 Banner University of Arizona Medical Center; Michael J. Stephen; Drexel University
 College of Medicine / Drexel Adult Cystic Fibrosis Center; Jorge Lascano, UF Clinical
 Research Center; Bennie C. McWilliams, Austin Children's Chest Associates; Brian
 Morrissey, University of California Davis Medical Center; Arvey Stone, Advocate
 Children's Hospital - Park Ridge / North Suburban Pulmonary and Critical Care
 Consultants; James Wallace, Sanford Research / USD; Jamie Wooldridge, St. Louis
 University; Stéphanie Bui, Groupe Hospitalier Pellegrin, CHU De Bordeaux; Donatello
 Salvatore, Centro Regionale Fibrosi Cistica, A.O. Ospedale San Carlo; Chantal
 Belleguic, Hôpital Pontchaillou CHU de Rennes; Lea Bentur, Pediatric Pulmonary Unit
 Rambam Medical Center; Ori Efrati, Sheba Medical Center; Eitan Kerem, Hadassah
 Medical Organization; Huda Mussaffi-Georgy, Schneider Children's Medical Center;
 Peter Barry, Wythenshawe Hospital; Diana Elizabeth Tullis, St. Michael's Hospital;
 Bradley Quon, St. Paul's Hospital; Larry C. Lands, McGill University Health Centre, Glen
 Site, Montreal Children's Hospital; Okan Elidemir, Sacred Heart Hospital; Felix
 Ringshausen, Hannover Medical School; Matthias Giese, Dr. von Haunersches
 Kinderspital; Nico Derichs, Charite Paediatric Pulmonology Department;
 Sivagurunathan Sutharsan, Ruhrlandklinik; Rainald Fischer, Pneumologische Praxis

Pasing; Jane C. Davies, Royal Brompton & Harefield NHS Foundation Trust, Royal Brompton Hospital; Gordon MacGregor, Queen Elizabeth University Hospital; Nicholas Withers, Royal Devon and Exeter NHS Foundation Trust, Royal Devon and Exeter Hospital; Daniel Gavin Peckham, The Leeds Teaching Hospitals NHS Trust, St. James University Hospital; Martin James Ledson, Liverpool Heart and Chest Hospital; Dominique Hubert, Hopital Cochin; Isabelle Semet-Gaudelus, Hopital Necker, Enfants Malades; Marleen Bakker, Erasmus Medical Center; Kors van der Ent, University Medical Center, Utrecht, Department of Pulmonology and Tuberculosis; Raphael Chiron, Hopital Arnaud de Villeneuve; Philippe Reix, CHU Lyon - Hopital Femme Mere-Enfant; Eva Van Braeckel, Universitair Ziekenhuis Gent; Nadine Desmazes-Dufeu, CHU Marseille - Hopital Nord; Anne Prévotat, CHRU de Lille - Hopital Albert Calmette; Harry Heijerman, HagaZiekenhuis; Elisabeth Josephien Maria Weersink, Academic Medical Center; Peter Cooper, The Children's Hospital at Westmead; Simon David Bowler, Mater Adult Hospital; Peter Gordon Middleton, Westmead Hospital; John William Wilson, The Alfred Hospital; Hugh Greville, Royal Adelaide Hospital; Carlo Castellani, Azienda Ospedaliera di Verona-Ospedale Civile Maggiore; Carla Colombo, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico; Vincenzina Lucidi, Ospedale Pediatrico Bambino Gesù, Centro Fibrosi Cistica; Serena Quattrucci, Umberto I Policlinico di Roma Università di Roma La Sapienza; Benedetta Fabrizzi, Centro di Riferimento Regionale per la Diagnosi Ancona e la Cura della Fibrosi Cistic.

SUPPLEMENTAL METHODS

Criteria for Eligible Residual Function Mutations

Population-level clinical criteria for residual function mutations were an average sweat chloride level <86 mmol/L and incidence of pancreatic insufficiency ≤50%. Laboratory criteria for residual function were the presence of mature CFTR by Western blot and observed chloride transport in the absence of treatment in isogenic cell lines expressing the *CFTR* gene in question.¹ In vitro response to ivacaftor was defined by a statistically significant increase in chloride transport in the presence of ivacaftor compared to baseline and/or an increase in chloride transport of ≥ 10% over baseline. A list of eligible mutations leading to residual function are given in Table S1.

Prespecified Patient-Level Eligibility Criteria

If the sweat chloride value was <60 mmol/L, evidence of chronic sinopulmonary disease was required, including at least 1 of the following: persistent colonization/infection with typical cystic fibrosis pathogens, including *Staphylococcus aureus*, *Haemophilus influenzae*, and mucoid and nonmucoid *Pseudomonas aeruginosa*; chronic cough and sputum production; persistent chest radiograph abnormalities (e.g., bronchiectasis, atelectasis, infiltrates, hyperinflation); nasal polyps, chronic sinusitis; and radiographic or computed tomographic abnormalities of the paranasal sinuses.

Sample Size and Power

The null hypothesis tested was that the mean absolute change from study baseline in percentage of predicted forced expiratory volume in 1 second (FEV₁) to the average of the week 4 and week 8 measurements was the same for (i) tezacaftor-ivacaftor and placebo and (ii) ivacaftor monotherapy and placebo. The sample size of 34 patients per sequence was adequate to yield at least 90% power to detect a treatment difference of 3 percentage points between tezacaftor-ivacaftor and placebo comparing the mean values of the primary endpoint; assuming a drop-out rate of 10%. A standard deviation (SD) of 7 percentage points and 2-sided significance level of 0.05 was used in the sample size calculations. Accounting for the testing strategy, the proposed sample size yielded approximately an 85% chance of observing a statistically significant difference between ivacaftor monotherapy and placebo for the primary endpoint, under the assumption that ivacaftor monotherapy was also 3 percentage points better than placebo.

Analysis of Primary Endpoint Variables

The primary analysis model included the absolute change from study baseline in percentage of predicted FEV₁ to the average of the week 4 and week 8 measurements as the dependent variable and the following fixed effects: treatment, period, percentage of predicted FEV₁ at study baseline, and patient as a random effect. The within-patient covariance was assumed to have the same compound symmetry (CS) structure for sequences containing placebo treatment but was different from the CS structure for sequences containing active treatment in both periods. Denominator degrees of freedom for the F-test for fixed effects was estimated using the Kenward-Roger

approximation.² The estimated mean of the dependent variable, a 95% confidence interval (CI), and a 2-sided P value were provided for each treatment. Similarly, the estimated between-group treatment differences along with the corresponding 95% CI and 2-sided P values were presented. No imputation of missing data were performed. Patients who had data only for one of the periods had a data structure similar to a parallel-group trial. Assuming that these patients had dropped out at random, the mixed effects model combined the estimate of treatment effect based on such patients with the estimate from patients who had data in both treatment periods. The weights used for combining these two estimates is based on their precisions.

Mixed-Model for Repeated Measures Analysis of Primary Endpoint

An alternative approach to analyze the absolute change from study baseline in percentage of predicted FEV₁ was to use the mixed-model for repeated measures (MMRM) approach. In the MMRM analysis, the absolute change from study baseline at each postbaseline visit (day 15, week 4, and week 8) during each treatment period was the dependent variable. The fixed effects in the model were: treatment, period, visit within period, treatment-by-visit interaction, and percentage of predicted FEV₁ at study baseline. The within-patient covariance was assumed to be unstructured (UN) for levels of period and UN for visits within period. The direct product of the 2 produced the estimated covariance matrix (type = UN@UN in SAS Procedure Mixed). The denominator degrees of freedom for the F-test for fixed effects was estimated using the Kenward-Roger approximation.² The average change from study baseline in percentage of predicted FEV₁ at weeks 4 and 8 for each treatment was estimated using contrasts from the MMRM. The estimated difference between treatments was also estimated

similarly. The resultant estimates, the 95% CI, and 2-sided P value were presented. A similar approach was followed to present the estimates for each visit. Additionally, the estimated change at each visit and the 95% CI were plotted.

Subgroup Analysis of the Primary Efficacy Endpoint

The subgroup analyses of the primary endpoint were performed using a model similar to that for the primary analysis. The primary result obtained from the model was the estimated difference between the treatment groups. The following subgroups were considered:

- Age at screening (<18 and ≥18 years)
- Percentage of predicted FEV₁ at study baseline (<40, ≥40 to <70, and ≥70)
- Residual function mutation type (CFTR class V noncanonical splice vs. CFTR classes II to IV residual function)
- Sex
- Region (North America and Europe [including Israel and Australia])
- Use of inhaled antibiotic (Yes, No)
- Use of inhaled bronchodilator (Yes, No)
- Use of inhaled hypertonic saline (Yes, No)
- Use of inhaled corticosteroids (Yes, No)
- Use of azithromycin (Yes, No)
- Colonization of *Pseudomonas aeruginosa* (Positive, Negative)

Each of the above subgrouping factors utilized a model analogous to the one used for the primary analysis but included an additional covariate for the relevant grouping factor

as well as a term for interaction with treatment. For the subgroup analysis on percentage of predicted FEV₁ severity at study baseline, the term percentage of predicted FEV₁ at study baseline was removed from the primary analysis model to avoid redundancy. For each subgroup, the estimated mean of the primary endpoint, the corresponding 95% CI and 2-sided P value are presented by treatment group. Similarly, the estimated between-group treatment differences along with the corresponding 95% CI and 2-sided P values are presented. The estimated between-group treatment differences in different subgroup categories were presented in a forest plot.

Analysis of Key Secondary Efficacy Endpoint

Analysis for the absolute change from study baseline in Cystic Fibrosis Questionnaire-Revised (CFQ-R) respiratory domain (pooled 'Children Ages 12 and 13' version and 'Adolescents and Adults' version) score to the average of week 4 and week 8 scores in each treatment period was similar to the primary analysis of the primary efficacy endpoint. However, percentage of predicted FEV₁ at study baseline was replaced by CFQ-R respiratory domain score at study baseline in the model.

Other Secondary Endpoints

Other secondary endpoints include (i) relative change in percentage of predicted FEV₁ and (ii) absolute change in sweat chloride from study baseline to the average of the week 4 and week 8 measurements in each treatment period. The analysis was similar to the primary analysis of the primary efficacy endpoint. However, for the analysis of

absolute change in sweat chloride the percentage of predicted FEV₁ study baseline was replaced by sweat chloride at study baseline in the model.

Multiplicity Adjustment

The testing strategy considered the comparison of tezacaftor-ivacaftor versus placebo and ivacaftor monotherapy versus placebo. To control for multiplicity of endpoints and treatments (the probability of Type 1 error), each endpoint was to be assessed sequentially using a gatekeeping approach where statistical significance could be claimed for the key secondary endpoint only if the primary endpoint met the requirements for significance. Additionally, as there were two treatment comparisons for each endpoint, the gatekeeping approach was applied (i.e., ivacaftor monotherapy for a given endpoint could achieve significance only if the comparison for tezacaftor-ivacaftor for the same endpoint was significant). For each endpoint, and for each treatment group, the comparison with placebo was to be conducted using a significance level (alpha) set at 0.05 (2-sided).

SUPPLEMENTAL RESULTS

Exploratory and Additional Supportive Endpoints

The mean absolute change from study baseline in body mass index (BMI) at week 8 was 0.34 kg/m² for tezacaftor-ivacaftor, 0.47 kg/m² for ivacaftor, and 0.18 kg/m² for placebo. The short treatment duration may have affected sensitivity to detect changes in BMI.

Although the study was not powered to evaluate changes in the rate of pulmonary exacerbation, the estimated event rate of pulmonary exacerbation was lower for tezacaftor-ivacaftor (0.34 events per year) and ivacaftor (0.29 events per year) than for placebo (0.63 events per year), noting these changes did not reach statistical significance. Compared with placebo, the rate ratio was 0.54 (95% CI: 0.26, 1.13) for tezacaftor-ivacaftor and 0.46 (95% CI: 0.21, 1.01) for ivacaftor (Table 2). The rate ratio was 1.18 (95% CI: 0.49, 2.87) for tezacaftor-ivacaftor compared with ivacaftor (Table 2).

Treatment with tezacaftor-ivacaftor and ivacaftor resulted in reduction of mean immunoreactive trypsinogen levels by day 15 that were sustained through week 8. The within-group mean change in immunoreactive trypsinogen from study baseline to week 8 was -18.1 ng/mL in the tezacaftor-ivacaftor group, -23.2 ng/mL in the ivacaftor group, and -2.1 ng/mL in the placebo group (Table 2).

At study baseline, the mean fecal elastase-1 value was 412.4 µg/g in the tezacaftor-ivacaftor group, 405.8 µg/g in the ivacaftor group, and 414.4 µg/g in the placebo group; a total of 22 (13.7%) patients in the tezacaftor-ivacaftor group, 22 (14.1%) patients in the ivacaftor group, and 21 (13.0%) patients in the placebo group had <200 µg/g of fecal elastase-1 (pancreatic insufficiency) at study baseline, reflecting a high prevalence of pancreatic sufficiency at baseline. The within-group mean change in fecal elastase-1 from study baseline to the average of week 4 and week 8 was -3.4 µg/g in the tezacaftor-ivacaftor group, -16.1 µg/g in the ivacaftor group, and -23.1 µg/g in the placebo group (Table 2). Among patients with values <200 µg/g at study baseline, 6 (27.2%) patients in the tezacaftor-ivacaftor group, 4 (18.2%) patients in the ivacaftor

group, and 1 (4.8%) patient in the placebo group had maximum fecal elastase-1 values ≥ 200 $\mu\text{g/g}$ at week 4 and week 8.

Safety

Additional Information on Adverse Events

Serious and life-threatening adverse events are reported in the main text. Serious adverse events, a subset of the above, occurred in 8 (4.9%) patients in the tezacaftor-ivacaftor group, 10 (6.4%) patients in the ivacaftor group, and 14 (8.6%) patients in the placebo group (Table 3).

Adverse events that led to treatment discontinuation are reported in the main text.

Adverse events led to treatment interruption in 2 (1.2%) patients in the tezacaftor-ivacaftor group, 5 (3.2%) patients in the ivacaftor group, and 6 (3.7%) patients in the placebo group.

Respiratory Adverse Events

Increased monitoring was conducted for respiratory adverse events because of the increased prevalence of acute but transient respiratory adverse events associated with the CFTR corrector lumacaftor.³⁻⁵ Adverse events associated with respiratory events occurred in 14 (8.6%) patients in the tezacaftor-ivacaftor group, 7 (4.5%) patients in the ivacaftor group, and 22 (13.6%) patients in the placebo group (Table S2 in the Supplementary Appendix). Overall and by preferred term, most adverse events associated with respiratory events either occurred at a similar incidence between the 3 treatment groups or were more common in the placebo group than in the ivacaftor or tezacaftor-ivacaftor groups. Respiratory events were mild to moderate in severity across

all treatment groups, and there were no grade 3 or 4 respiratory events. There were no respiratory events that were serious or led to death.

Adverse events associated with respiratory symptoms (chest discomfort, dyspnea, and respiration abnormal), occurred in 11 (6.8%) patients in the tezacaftor-ivacaftor group, 3 (1.9%) patients in the ivacaftor group, and 16 (9.9%) patients in the placebo group. The events either occurred at a similar incidence between the 3 treatment groups or were more common in the placebo group than in the ivacaftor or tezacaftor-ivacaftor groups. The time-to-onset of the first adverse event associated with respiratory events and symptoms was similar in all 3 treatment groups.

Acute Effects on Spirometry

Postdose spirometry assessments were performed for patients (n=21, 20, and 24 for tezacaftor-ivacaftor, ivacaftor and placebo, respectively) between the ages of 12 and 18 years. The postdose (2- and 4-hour) percentage of predicted FEV₁ values showed no evidence of acute decline from the predose values on both days 1 and 15 for either tezacaftor-ivacaftor or ivacaftor (Table S3 in the Supplementary Appendix).

Liver Function Tests

Few patients had elevations in liver transaminases or total bilirubin during the study period (Table S4), and no patients experienced an elevated transaminase > 3 x ULN concurrent with an elevated total bilirubin > 2 x ULN. One (0.6%) patient in the placebo group, 3 (1.9%) patients in the ivacaftor group, and 1 (0.6%) patient in the tezacaftor-ivacaftor group had transaminase elevations >3 x upper limit of normal (ULN). Two

(1.3%) patients in the ivacaftor group had transaminase elevations $>5 \times$ ULN. No patient had transaminase elevations $>8 \times$ ULN during the treatment period. One (0.6%) patient in the placebo group, 2 (1.3%) patients in the ivacaftor group, and 2 (1.2%) patients in the tezacaftor-ivacaftor group had total bilirubin elevations $>2 \times$ ULN. None of these were serious or led to treatment discontinuation, and the majority were assessed as mild or moderate by the investigator.

SUPPLEMENTAL DISCUSSION

Residual pancreatic function allows for the potential to preserve organ function or delay the onset of pancreatic insufficiency with early CFTR modulation treatment. Fecal elastase-1 did not significantly change and the majority of subjects met the criteria for pancreatic sufficiency at baseline (FE-1 $>200 \mu\text{g/g}$); in contrast, serum immunoreactive trypsinogen suggested active treatment may improve pancreatic injury in patients with residual pancreatic secretion. Similar immunoreactive trypsinogen responses were seen with ivacaftor in 2- to 5-year old patients with cystic fibrosis carrying the *G551D* allele.⁶ Further study of the potential for pancreatic organ preservation is warranted, targeting patients early in life, before organ damage is less reversible. Similarly, the potential to reduce pulmonary exacerbations and increase BMI over long-term treatment requires further study and may raise unique issues in patients with residual function since exacerbations can be less frequent in this population.

SUPPLEMENTAL REFERENCES

1. Yu YC, Sohma Y, Hwang TC. On the mechanism of gating defects caused by the R117H mutation in cystic fibrosis transmembrane conductance regulator. *J Physiol* 2016;594:3227-44.
2. Kenward MG, Roger JH. Small sample inference for fixed effects from restricted maximum likelihood. *Biometrics* 1997;53:983-97.
3. Wainwright CE, Elborn JS, Ramsey BW, et al. Lumacaftor-ivacaftor in patients with cystic fibrosis homozygous for Phe508del CFTR. *N Engl J Med* 2015;373:220-31.
4. Milla CE, Ratjen F, Marigowda G, et al. Lumacaftor/ivacaftor in patients aged 6-11 years with cystic fibrosis and homozygous for F508del-CFTR. *Am J Respir Crit Care Med* 2017;195:912-20.
5. Ratjen F, Hug C, Marigowda G, et al. Efficacy and safety of lumacaftor and ivacaftor in patients aged 6-11 years with cystic fibrosis homozygous for F508del-CFTR: a randomised, placebo-controlled phase 3 trial. *Lancet Respir Med* 2017 Jun 8 [Epub ahead of print].
6. Davies JC, Cunningham S, Harris WT, et al. Safety, pharmacokinetics, and pharmacodynamics of ivacaftor in patients aged 2-5 years with cystic fibrosis and a CFTR gating mutation (KIWI): an open-label, single-arm study. *Lancet Respir Med* 2016;4:107-15.

SUPPLEMENTAL TABLES

Table S1. Eligible Residual Function Mutations

<i>Noncanonical splice mutations</i>	no.
2789+5G→A	37
3849+10kbC→T	69
3272-26A→G	36
711+3A→G	3
<i>Missense mutations</i>	
E56K	0
P67L	17
E831X	1
R74W	0
D110E	0
D110H	1
R117C	1
E193K	0
L206W	5
R347H	4
R352Q	3
R1070W	3
A455E	20
F1074L	0
D579G	3
D1152H	26
S945L	13
D1270N	0
S977F	2
F1052V	0
K1060T	0

Table S2. Summary of Treatment-Emergent Respiratory Events by Preferred Term, Safety Set.

	Placebo N=162 no. (%)	Ivacaftor N=157 no. (%)	Tezacaftor- Ivacaftor N=162 no. (%)
Any AEs (respiratory events)	22 (13.6)	7 (4.5)	14 (8.6)
Chest discomfort	0	0	2 (1.2)
Dyspnea	11 (6.8)	3 (1.9)	9 (5.6)
Respiration abnormal	5 (3.1)	0	3 (1.9)
Asthma	3 (1.9)	0	0
Bronchial hyperreactivity	0	0	0
Bronchospasm	2 (1.2)	0	0
Wheezing	3 (1.9)	4 (2.5)	3 (1.9)
Any respiratory events by maximum severity	22 (13.6)	7 (4.5)	14 (8.6)
Mild	12 (7.4)	5 (3.2)	12 (7.4)
Moderate	10 (6.2)	2 (1.3)	2 (1.2)
Severe	0	0	0
Life-threatening	0	0	0
Missing	0	0	0
Events leading to treatment discontinuation	1 (0.6)	0	0
Serious events	0	0	0
Related serious events	0	0	0
Events leading to death	0	0	0
By time interval			
Patients with any respiratory events			
>0 to ≤1 week	5 (3.1)	1 (0.6)	4 (2.5)
>1 to ≤2 weeks	1 (0.6)	1 (0.6)	1 (0.6)
>0 to ≤8 weeks	15 (9.3)	5 (3.2)	9 (5.6)
>8 weeks	7 (4.3)	2 (1.3)	7 (4.3)

AEs, adverse events; MedDRA, Medical Dictionary for Regulatory Activities.

Note: Respiratory events were coded using MedDRA Version 19.1. If a patient had multiple events within a category, system organ class, or preferred term, the patient was counted only once. Related serious events include related, possibly related, and missing categories.

Table S3. Mean (SD) Change from Predose to Post Dose in Percentage of Predicted FEV₁ on Days 1 and 15, Safety Set for Patients ≥ 12 to <18 Years Old at Screening.

	Placebo N=24	Ivacaftor N=20	Tezacaftor- Ivacaftor N=21
Day 1			
no.	12	12	13
2 hours post dose	-0.5 (3.9)	0.9 (5.7)	0.5 (3.6)
no.	12	13	14
4 hours post dose	-0.3 (4.8)	-0.4 (6.8)	0.9 (3.9)
Day 15			
no.	13	12	12
2 hours post dose	1.3 (3.6)	1.9 (2.4)	1.4 (2.5)
no.	13	12	10
4 hours post dose	0.7 (4.1)	2.9 (5.1)	1.8 (3.4)

FEV₁, forced expiratory volume in 1 second; no., size of subsample; N, total sample size; SD, standard deviation.

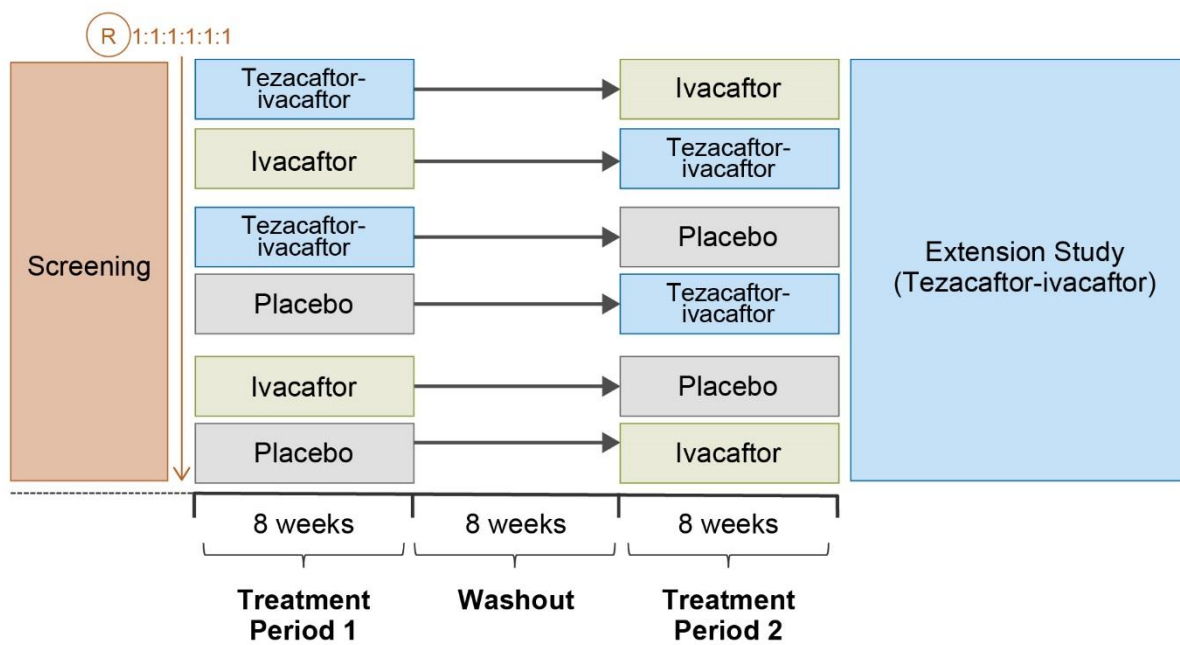
Table S4. Liver Transaminase and Bilirubin Elevations During the Study Period.

Parameter	Placebo N=162	Ivacaftor N=157	Tezacaftor- Ivacaftor N=162
Alanine aminotransferase — n/N1 (%)			
>ULN to ≤3x ULN	12/162 (7.4)	18/157 (11.5)	18/162 (11.1)
>3x to ≤5x ULN	1/162 (0.6)	3/157 (1.9)	1/162 (0.6)
>5x ULN	0/162	0/157	0/162
Aspartate aminotransferase — n/N1 (%)			
>ULN to ≤3x ULN	17/162 (10.5)	21/157 (13.4)	23/162 (14.2)
>3x to ≤5x ULN	0/162	2/157 (1.3)	1/162 (0.6)
>5x ULN	0/162	2/157 (1.3)	0/162
Total bilirubin — n/N1 (%)			
>1.5x ULN to ≤2x ULN	0/162	3/157 (1.9)	2/162 (1.2)
>2x ULN to ≤3x ULN	1/162 (0.6)	2/157 (1.3)	2/162 (1.2)
>3x ULN	0/162	0/157	0/162

ULN, upper limit of normal.

SUPPLEMENTAL FIGURES

Figure S1. Study Design.



R, randomized.

Figure S2. CONSORT Diagram.

